

Glycosyl hydrolase genes and their use for producing enzymes for the biodegradation of carrageenans

The present invention relates to glycosyl hydrolase genes for the biotechnological production of oligosaccharides, especially sulfated oligo-carrageenans and more particularly oligo-iota-carrageenans and oligo-kappa-carrageenans, by the biodegradation of carrageenans.

The sulfated galactans of Rhodophyceae, such as agars and carrageenans, represent the major polysaccharides of Rhodophyceae and are very widely used as gelling agents or thickeners in various branches of activity, especially agri-foodstuffs. About 6000 tonnes of agars and 22,000 tonnes of carrageenans are extracted annually from red seaweeds for this purpose. Agars are commercially produced by red seaweeds of the genera *Gelidium* and *Gracilaria*. Carrageenans, on the other hand, are widely extracted from the genera *Chondrus*, *Gigartina* and *Eucheuma*.

Carrageenans consist of repeat D-galactose units alternately bonded by β 1 \rightarrow 4 and α 1 \rightarrow 3 linkages. Depending on the number and position of sulfate ester groups on the repeat disaccharide of the molecule, carrageenans are thus divided into several different types, namely: kappa-carrageenans, which possess one sulfate ester group, iota-carrageenans, which possess two sulfate ester groups, and lambda-carrageenans, which possess three sulfate ester groups.

The physicochemical properties and the uses of these polysaccharides as gelling agents are based on their capacity to undergo ball-helix conformational transitions as a function of the thermal and ionic environment [Kloareg et al., Oceanography and Marine Biology - An annual review 26 : 259-315 (1988)].

Furthermore, carrageenans are structural analogs of the sulfated polysaccharides of the animal extracellular matrix (heparin, chondroitin, keratan, dermatan) and they exhibit biological activities which are related to certain functions of these glycosaminoglycans.

In particular, carrageenans are known:

- (i) - for their action on the immune system, causing the secretion of interleukin or prostaglandins,
- (ii) - for their antiviral action on the AIDS virus HIV1, the herpes virus HSV1 and the hepatitis A virus,

- (iii) - as antagonists of the fixation of the growth factors of human cells,
- (iv) - and also for their action on the proliferation of keratinocytes and their action on the contractility of fibroblasts.

Furthermore, oligocarrageenans act on the adherence, the division and the protein synthesis of human cell cultures, doubtless as structural analogs of the glycosylated part of the proteins of the extracellular matrix. In plants, oligocarrageenans very significantly elicit enzymatic activities which are markers of growth (amylase) or of the phenolic defense metabolism (laminarinase, phenyl-alanineammonium lyase).

Carrageenans are extracted from red seaweeds by conventional processes such as hot aqueous extraction, and oligocarrageenans are obtained from carrageenans by chemical hydrolysis or, preferably, by enzymatic hydrolysis.

The production of oligocarrageenans by enzymatic hydrolysis generally comprises the following steps:

- 1) production of a glycosyl hydrolase by the culture of a marine bacterium;
- 2) enzymatic hydrolysis of the carrageenan with the glycosyl hydrolase thus obtained; and
- 3) fractionation and purification of the oligocarrageenans obtained.

Microorganisms which produce enzymes capable of hydrolyzing iota- and kappa-carrageenans were isolated by Bellion et al. in 1982 [Can. J. Microbiol. **28** : 874-80 (1982)]. Some are specific for κ - or ι -carrageenan and others are capable of hydrolyzing both substrates. Another group of bacteria capable of degrading carrageenans was characterized by Sarwar et al. in 1983 [J. Gen. Appl. Microbiol. **29** : 145-55 (1983)]. These yellow-orange bacteria are assigned to the *Cytophaga* group of bacteria and some of these bacteria have the property of hydrolyzing both agar and carrageenans.

Purification and characterisation of several ι -carrageenases and κ -carrageenases, such as the ι -carrageenase and κ -carrageenase of *Cytophaga drobachiensis*, the ι -carrageenase of *Alteromonas fortis* and the κ -carrageenase of *Alteromonas carrageenovora*, were described in the thesis of P. Potin ["Recherche, production, purification et caractérisation de galactane-hydrolases pour la préparation des parois d'algues rouges", (February 1992)]. A detailed study of the κ -carrageenase of *Alteromonas carrageenovora* was described by Potin et al. [Eur. J. Biochem. **228**, 971-975 (1995)].

The availability of specific enzymes and tools for obtaining oligocarrageenans by genetic engineering could markedly improve their production.

The Applicant has now found novel glycosyl hydrolase genes which make it possible specifically to obtain either oligo-iota-carrageenans or oligo-kappa-carrageenans.

Thus the present invention relates to novel genes which code for glycosyl hydrolases having an HCA score with the iota-carrageenase of *Alteromonas fortis* which is greater than or equal to 65%, preferably greater than or equal to 70% and advantageously greater than or equal to 75% over the domain extending between amino acids 164 and 311 of the sequence [SEQ ID No. 2] of the iota-carrageenase of *Alteromonas fortis*.

The present invention relates more particularly to the nucleic acid sequence [SEQ ID No. 1] which codes for an iota-carrageenase as defined above, the amino acid sequence of which is the sequence [SEQ ID No. 2].

The present invention further relates to the genes which code for glycosyl hydrolases having an HCA score with the kappa-carrageenase of *Alteromonas carrageenovora* which is greater than or equal to 75%, preferably greater than 80% and advantageously greater than 85% over the domain extending between amino acids 117 and 262 of the sequence [SEQ ID No. 6] of the kappa-carrageenase of *Alteromonas carrageenovora*.

In particular, the invention relates to the nucleic acid sequence [SEQ ID No. 7] which codes for a kappa-carrageenase having a score as defined above, the amino acid sequence of which is the sequence [SEQ ID No. 8].

The glycosyl hydrolase genes of the invention are obtained by a process which consists in selecting proteins having an HCA score with the iota-carrageenase of *Alteromonas fortis* which is greater than or equal to 65%, preferably greater than or equal to 70% and advantageously greater than or equal to 75% over the domain extending between amino acids 164 and 311 of the sequence [SEQ ID No. 2] of the iota-carrageenase of *Alteromonas fortis*, and in sequencing the resulting genes by the conventional techniques well known to those skilled in the art.

The glycosyl hydrolase genes of the invention can also be obtained by a process which consists in selecting proteins having an HCA score with the kappa-carrageenase of *Alteromonas carrageenovora* which is greater than or equal to 75%, preferably greater than 80% and advantageously greater than 85% over the domain extending between amino acids 117 and 262 of the sequence [SEQ ID No. 6] of the kappa-carrageenase of *Alteromonas carrageenovora*, and in

sequencing the resulting genes by the conventional techniques well known to those skilled in the art.

Finally, the present invention relates to the use of the above glycosyl hydrolase genes for obtaining, by genetic engineering, glycosyl hydrolases which are useful for the biotechnological production of oligocarrageenans.

The glycosyl hydrolases according to the invention are therefore characterized by the HCA score which they possess with a particular domain of the amino acid sequence of the iota-carrageenase of *Alteromonas fortis* or the kappa-carrageenase of *Alteromonas carrageenovora*.

The HCA or "Hydrophobic Cluster Analysis" method is a method of analyzing the sequences of proteins represented as a two-dimensional structure, which has been described by Gaboriaud et al. [FEBS Letters 224, 149-155 (1987)].

It is known that the three-dimensional structure of a protein governs its biological properties, the production of an active protein demanding correct folding.

It is also known that the primary structure of proteins varies much more substantially than the higher-order structures and that proteins can be grouped into families which show similar secondary and tertiary structures but sometimes have such divergent primary sequences that the mutual relationship between such proteins is not obvious. The code which relates primary structure and secondary structure therefore appears to be highly degenerate since very different primary structures can ultimately lead to similar secondary and tertiary structures [Structure 3, 853-859 (1995) and Proc. Natl. Acad. Sci. USA 92 (1995)].

The use of the HCA method has shown that the distribution, size and shape of these hydrophobic clusters along the amino acid sequences are representative of the 3D folding of the proteins studied.

Also, Woodcock et al. [Protein Eng. 5, 629-635 (1992)] have shown that the hydrophobic clusters defined by the α -helical 2D diagram are statistically centered on the regular secondary structures (α -helices, β -strands), that the 2D diagram based on the α -helix carries the greatest amount of structural information and that the correspondence between hydrophobic clusters and elements of secondary structure is of the same quality for any type of folding (all α , all β , α/β and $\alpha + \beta$), thus demonstrating that the HCA method can be used irrespective of the type of protein.

L. Lemesle-Varloot et al. [Biochimie 72, 555-574 (1990)] have shown that when two proteins have a similar distribution of hydrophobic clusters over a domain of at least 50 residues, their three-dimensional structures in this domain are considered to be superimposable and their functions to be analogous.

Thus, for example, Barbeyron et al. [Gene 139, 105-109 (1994)] used this HCA method for the comparison of the similarities in the shape, distribution and size of several hydrophobic clusters of the κ -carrageenase of *Alteromonas carrageenovora* with respect to enzymes from family 16 of glycosyl hydrolases.

The two-dimensional representation used in the HCA method is an α -helix in which the amino acids are arranged by computer processing to give 3.6 residues per turn. To obtain an easily readable plane image, the helix is cut in the longitudinal direction. Finally, to obtain the whole of the hydrophobic clusters situated at the edges of the image, the diagram is duplicated. The method uses a code which recognizes only two states: the hydrophobic state and the hydrophilic state.

The amino acids recognized as being hydrophobic are identified and grouped into characteristic geometric figures. Using these two states makes it possible to become independent of the tolerance shown by the two- and three-dimensional structures towards the variability of the primary sequences. Furthermore, this representation affords rapid observation of interactions over a short or medium distance since the first amino acid and the second, adjacent amino acid of a given residue are located on a segment of 17 amino acids. Finally, in contrast to the analytical methods based on the primary or secondary structures of proteins, no "window" of predefined length is used.

The fundamental characteristic of the α -helix representation is that, for a given globular protein or only a domain of this protein, the distribution of the hydrophobic residues on the diagram is not random. The hydrophobic residues (VILFWMY) form clusters of varying geometry and size. On the diagram, the hydrophilic and hydrophobic faces of the amphiphilic helices are very recognizable. Thus a horizontal diamond cluster corresponds to the hydrophobic face of an α -helix, the internal helices appear as large horizontal hydrophobic clusters and the β -strands appear as rather short, vertical hydrophobic clusters. The method makes it possible to identify the hydrophobic residues forming the core of the globular proteins and to locate the elements of secondary structure, namely the α -helices and the β -strands, independently of any knowledge of the secondary structure of the protein studied.

The HCA score between two proteins is calculated as follows:

For each cluster:

$$\text{HCA score} = 2\text{CR}/(\text{RC}_1 + \text{RC}_2) \times 100\%$$

where

- RC_1 and RC_2 are the number of hydrophobic residues in the cluster of protein 1 (cluster 1) and the cluster of protein 2 (cluster 2), respectively.

- CR is the number of hydrophobic residues in the cluster 1 which correspond to the hydrophobic residues in the cluster 2.

The mean value obtained for all the clusters along the protein sequences compared gives the final HCA score.

On the HCA profiles, the amino acids are represented by their standard code of a single letter, with the exception of proline (P), glycine (G), serine (S) and threonine (T).

In fact, because of their particular properties, these residues are represented by the special symbols indicated below so as to facilitate their visual identification on the HCA diagrams (cf. list of abbreviations).

Proline introduces high constraints into the polypeptide chain and is considered systematically as an interruption in the clusters. In fact, proline residues stop or deform the helices and the lamellae. Glycine possesses a very substantial conformational flexibility because of the absence of a side chain in this amino acid. Serine and threonine are normally hydrophilic, but they can also be found in hydrophobic environments, such as α -helices, in which their hydroxyl group loses their hydrophilic character because of the hydrogen bond formed with the carbonyl group of the main chain. Within the hydrophobic β -lamellae, threonine is sometimes capable of replacing hydrophobic residues by virtue of the methyl group on its side chain.

Amino acids can be divided into four groups according to their hydrophobicity:

(i) - strongly hydrophobic residues: V, I, L and F;

(ii) - moderately hydrophobic residues: W, M and Y

→ W appears at surface sites more frequently than F,

→ M is encountered at various sites, internal or otherwise,

→ Y can adapt to internal hydrophobic environments and is frequently found in loops;

(iii) - weakly hydrophobic residues: A and C are virtually insensitive to the hydrophobic character of their environment; and

(iv) - hydrophilic residues: D, E, N, Q, H, K and R.

Using this HCA method, the Applicant has found that proteins having an HCA score with the iota-carrageenase of *Alteromonas fortis* which is greater than or equal to 65% over the domain extending between amino acids 164 and 311 of said iota-carrageenase are enzymes of the glycosyl hydrolase type and more particularly iota-carrageenases appropriate for the production of oligo-iota-carrageenans from carrageenans.

The proteins having an HCA score which is greater than or equal to 70%, preferably greater than or equal to 75%, with the above domain 164-311 are particularly preferred for the purposes of the invention.

One particular example of glycosyl hydrolase obtained with a gene according to the invention is the protein having the amino acid sequence [SEQ ID No. 2], extracted from *Alteromonas fortis*.

Another particular example of glycosyl hydrolase obtained with a gene according to the invention is the protein having the amino acid sequence [SEQ ID No. 4], extracted from *Cytophaga drobachiensis*.

Likewise, the Applicant has found that proteins having an HCA score with the kappa-carrageenase of *Alteromonas carrageenovora* which is greater than or equal to 75% over the domain extending between amino acids 117 and 262 of said kappa-carrageenase are enzymes of the glycosyl hydrolase type and more particularly kappa-carrageenases appropriate for the production of oligo-kappa-carrageenans from carrageenans.

The proteins having an HCA score which is greater than or equal to 80%, preferably greater than or equal to 85%, with the above domain 117-262 are particularly preferred for the purposes of the invention.

The above proteins are advantageously extracted from marine bacteria.

One particular example of glycosyl hydrolase obtained with a gene according to the invention is the protein having the amino acid sequence [SEQ ID No. 6], extracted from *Alteromonas carrageenovora*.

Another particular example of glycosyl hydrolase obtained with a gene according to the invention is the protein having the amino acid sequence [SEQ ID No. 8], extracted from *Cytophaga drobachiensis*.

As indicated previously, the genes according to the invention, coding for glycosyl hydrolases, can be obtained by sequencing the genome of bacteria which product glycosyl hydrolases, as defined above, by the conventional methods well known to those skilled in the art.

The invention further relates to the expression vectors which carry the nucleic acid sequences according to the invention, with the means for their expression.

These expression vectors can be used to transform prokaryotic microorganisms, particularly *Escherichia coli*, or eukaryotic cells such as yeasts or fungi.

The invention will now be described in greater detail by means of the illustrative and non-limiting Examples below.

The methods used in these Examples are methods well known to those skilled in the art, which are described in detail in the work by Sambrook, Fristsch and Maniatis entitled "Molecular cloning: a laboratory manual", published in 1989 by Cold Spring Harbor Press, New York (2nd edition).

The following description will be understood more clearly with the aid of Figures 1 to 4, which respectively show the following:

Fig. 1: The maximum similarity alignment, according to the method of Needleman and Wunsch [J. Mol. Biol. 48, 443-453 (1970)], of the amino acid sequence of the iota-carrageenase of *Alteromonas fortis* (top part) and the iota-carrageenase of *C. drobachiensis* (bottom part).

Fig. 2: The HCA profiles of the amino acid sequences of the iota-carrageenases of *Cytophaga drobachiensis* and *Alteromonas fortis*.

Fig. 3: The maximum similarity alignment, according to the method of Needleman and Wunsch, 1970, J. Mol. Biol. 48, 443-453, of the amino acid sequence of the kappa-carrageenase of *Alteromonas carrageenovora* (top part) and *Cytophaga drobachiensis* (bottom part).

Fig. 4: The HCA profiles of the amino acid sequences of the kappa-carrageenases of *Cytophaga drobachiensis* and *Alteromonas fortis*.

The abbreviations or special symbols used for the amino acids in the Examples below are as follows:

	Glycine: \diamond
5	Proline: *
	Threonine : \square
	Sérine: $\square \cdot$
	Alanine: A
	Valine: V
10	Leucine: L
	Isoleucine: I
	Methionine: M
	Phenylalanine: F
	Tryptophan: W
15	Cysteine: C
	Asparagine: N
	Glutamine: Q
	Tyrosine: Y
	Aspartate: D
20	Glutamate: E
	Lysine: K
	Arginine: R
	Histidine: H

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SECTION 1: Cloning of the genes of the iota-carrageenases of *Cytophaga drobachiensis* and *Alteromonas fortis*

Genome libraries of the DNAs of *C. drobachensis* and *A. fortis* were constructed.

Ampicillin (50 µg/ml) or tetracycline (15 µg/ml) was added to the agar or non-agar culture media from stock solutions prepared in 50% ethanol (to avoid solidification at the storage temperature, -20°C), except in the case of the non-recombinant strain DH5α.

The total DNA of *C. drobachiensis* and the total DNA of *A. fortis* were prepared by the method described by Barbeyron et al. [J. Bacteriol. 160, 586-590 (1984)].

The purified DNA fragments of 5000 to 10,000 bp were cloned at the *Bam*HI site of plasmid pAT153, which cleaves the tetracycline resistance gene.

6000 clones were obtained in each of the genome libraries.

The five positive *C. drobachiensis* clones and the two positive *A. fortis* clones, which hollowed out a hole in the ι-carrageenan after one week of culture at 22°C, are referred to respectively as pIC1 to pIC5 and pIP1 to pIP2.

1. Cloning from *C. drobachiensis*

The cloning of this gene is described in detail by T. Barbeyron in the doctoral thesis examined on 28 October 1993 at the Université Pierre et Marie Curie, Roscoff.

The plasmid DNA was isolated from the above five clones by the alkaline lysis method [Nucleic Acid Res. 7 : 1513 (1979)].

The sizes and mapping of the inserts showing an ι-carrageenase activity were determined by agarose gel electrophoresis after single and double digestion of their plasmids with various restriction enzymes.

The DNA fragments were extracted from the agarose by the glass wool method.

All the plasmids obtained contain an identical *Pvu*II fragment of 3.3 kb.

This fragment was subcloned in phagemid pbluescript KSII (Stratagene) (pICP07 and pICP16).

Likewise, the internal *Nde*I fragment and a *Hind*III fragment partially comprising the *Pvu*II fragment were subcloned to give the pICN22 and pICH42 subclones, respectively.

To locate the ι-carrageenase gene, libraries were constructed from the pICP07 and pICP16 subclones in phagemid pbluescript with the aid of the exonuclease III of *E. coli*, using the "ExoIII" kit from Pharmacia.

The subclones and the ExoIII clones obtained were plated onto Zd medium solidified with ι-carrageenan.

Only the pICP16 and pICP07 clones and the ExoIII pICP074 and pICP0712 clones (obtained by degradation with ExoIII for 4 minutes and 12 minutes, respectively, from the pICP07 clone) are ι-carrageenase-positive.

2. Cloning from *Alteromonas fortis*

The DNA of the pIP1 and pIP2 clones showed inserts of 10.45 kb and 4.125 kb respectively, having a common fragment of 3 kb. These clones showed a positive ι-carrageenase activity. Different fragments were subcloned and plated as described above. However, none of the subclones obtained proved to be ι-carrageenase-positive.

SECTION 2: Determination of the nucleotide sequences of the genes coding for the ι -carrageenases of *Cytophaga drobachiensis* and *Alteromonas fortis*

1. Sequence of the *Cytophaga drobachiensis* gene

5 Plasmid pICP0712 was used to determine the nucleotide sequence of the gene responsible for the ι -carrageenase activity of *C. drobachiensis* [SEQ ID No. 3].

10 This nucleotide sequence is composed of 1837 bp. Translation of the six reading frames revealed only one open frame, called *cgiA*. The potential initiation codon is situated 333 bp beyond the 5'P end of the sequence.

15 The protein sequence [SEQ ID No. 4] deduced from the sequence of *cgiA* is composed of 391 amino acids, corresponding to a theoretical molecular weight of 53.4 kDa. The hydropathic profile of this protein shows a hydrophobic region covering the first 24 amino acids. The presence of a positively charged amino acid (Lys) followed by a hydrophobic block and then by a polar segment of six amino acids suggests that this domain could be a signal peptide. According to the analyses performed by the method of Von Heijne [J. Mol. Biol. 184 : 99-105 (1985)], the signal peptidase would cleave between valine (Val²⁴) and threonine (Thr²⁵). The mature protein devoid of its signal peptide would have a theoretical
20 molecular weight of 50.7 kDa. The identity of the *cgiA* gene was confirmed by determination of the amino acids at the NH₂ end of the partially purified protein. The sequence obtained matches the one deduced from the nucleotide sequence. The first amino acid is situated 14 residues from the NH₂ end generated by the signal peptidase. As the presence of the two prolines following the amino acids
25 determined by microsequencing had slightly disturbed the order of appearance of the N-terminal residues, the sequence of an internal oligopeptide, purified by HPLC after cleavage with trypsin, was established. The sequence NH₂ATYKCOOH obtained is situated near the C-terminal end of the ι -carrageenase (residues 396 to 399).

30 **2. Sequence of the *Alteromonas fortis* gene**

Plasmids pIHP15 and pIHPX17, subcloned from pIP1 and pIP2, were used to determine the nucleotide sequence of the gene responsible for the ι -carrageenase activity of *Alteromonas fortis*, SEQ ID No. 1. The 2085 bp fragment contains a single open reading frame of 1473 bp, called *cgiA*. The sequence situated upstream
35 of the initiation codon (ATG²¹¹) is not a coding sequence.

The protein sequence deduced from the sequence of the *A. fortis* ι-carrageenase gene [SEQ ID No. 2] consists of 491 amino acids, corresponding to a theoretical molecular weight of 54.802 kDa. In the present case, again, the N-terminal part of the protein exhibits a high hydrophobicity, suggesting that this domain could be a signal peptide; the hypothetical cleavage site would be situated between glycine (Gly²⁶) and alanine (Ala²⁷). The mature protein devoid of its signal peptide would have a theoretical molecular weight of 51.95 kDa, corresponding to a value similar to the molecular weight obtained with the protein purified by SDS-PAGE, namely 57 kDa.

SECTION 3: Comparison of the protein sequences of the ι-carrageenases of *Cytophaga drobachiensis* and *Alteromonas fortis*

After removal of the signal peptide from each sequence, it could be seen that the sequence of the ι-carrageenase of *C. drobachiensis* has similarities to that of the ι-carrageenase of *A. fortis*.

In fact, the two sequences of iota-carrageenase have a similarity of 43.2% over the whole of the linear sequence alignment. This similarity is particularly high (57.8%) between amino acids 164 and 311 (numbering of the iota-carrageenase of *Alteromonas fortis* (Fig. 1)).

At the same time, an HCA analysis showed that the HCA score between the two proteins is 82% over a domain of 293 amino acids and reaches 90.5% in the case of said domain 164-311 (Fig. 2).

No significant similarity to other polysaccharidases known hitherto could be demonstrated.

These two enzymes therefore constitute a novel family of glycosyl hydrolases.

EXAMPLE II:

The kappa-carrageenases of *Alteromonas carrageenovora* and *Cytophaga drobachiensis*

SECTION 1: Cloning of the kappa-carrageenase genes

Alteromonas carrageenovora ATCC 43555 was obtained from the American Type Culture Collection. The strains *A. carrageenovora* and *C. drobachiensis* were cultivated under conditions identical to those mentioned in section 1 of Example I.

Likewise, genome libraries were constructed using the strain *Escherichia coli* DH5α and plasmid vector pAT153.

1. Cloning from *Alteromonas carrageenovora*

The preparation of this gene is described in detail by T. Barbeyron in the thesis cited above (cf. Example 1) and in Gene 139, 105-109 (1994).

From the genome library of *Alteromonas carrageenovora*, 4 *E. coli* clones, called K1 to K4, were capable of hydrolyzing kappa-carrageenan.

Plasmids pKA1 to pKA4 were purified from the four independent clones and mapped with the aid of the restriction endonucleases *Bam*HI, *Dra*I, *Eco*RI, *Hind*III, *Mlu*I, *Pst*I, *Pvu*II, *Sal*I, *Ssp*I, *Xba*I and *Xho*I.

The presence of a 2.2 kb *Dra*I-*Hind*III fragment was noted in each plasmid.

This common fragment, which is the whole insert of plasmid pKA3, was sequenced in its entirety from plasmid pKA3.

2. Cloning from *Cytophaga drobachiensis*

From the genome library of *C. drobachiensis*, five *E. coli* clones, called pKC1 to pKC5, were capable of hollowing out a hole in the substrate. The plasmids isolated and purified from said clones were mapped with restriction endonucleases.

Internal fragments of 1100 bp and 600 bp respectively were subcloned from pKC1 in phagemid pbluescript and were called pKCE11 and pKCN6.

Plasmids pKC1, pKCE11 and pKCN6 were used to determine the nucleotide sequence of the kappa-carrageenase gene.

SECTION 2: Determination of the sequences of the genes coding for the kappa-carrageenases of *Alteromonas carrageenovora* and *Cytophaga drobachiensis*

1. Sequence of the *Alteromonas carrageenovora* gene

The number of nucleotides in the pKA3 insert is 2180 bp. Translation in the six reading frames reveals the presence of three open frames, only one of which is complete; this one separates the other two, which are only partial. All three of them are located on the same DNA strand. The second open frame, called cgkA, read in the third reading frame, contains 1191 bp [SEQ ID No. 5].

The translation product of the cgkA gene corresponds to a protein of 397 amino acids with a theoretical molecular weight of 44,212 Da (SEQ ID No. 6). The hydropathic profile of this protein shows a highly hydrophobic domain,

extending over 25 amino acids, at the N-terminal end. This domain comprises a positively charged amino acid (Lys) followed by a segment rich in hydrophobic amino acids and then by three polar amino acids. These results suggest that a signal peptide is involved. The N-terminal sequence of the protein purified from the culture supernatant was determined, thereby confirming the identity of the gene. These results indicate that the signal peptidase cleaves the protein between residues 25 and 26, which is consistent with Von Heijne's rule (-3, -1). The mature protein therefore has a theoretical molecular weight of 41.6 kDa.

2. Sequence of the *Cytophaga drobachiensis* gene

The pKC1 insert of 4425 bp contains a single open reading frame of 1635 bp, called *cgkA* (SEQ ID No. 7).

The protein translated from the kappa-carrageenase gene is a protein comprising 545 amino acids with a molecular weight of 61.466 kDa [SEQ ID No. 8].

The hydropathic profile of this protein shows a highly hydrophobic domain at the N-terminal end, suggesting that a signal peptide is involved.

According to Von Heijne's rule (-3, -1), the cleavage site of the signal peptidase should be situated between threonine and serine in positions 35 and 36 respectively, with the codon ATG⁸⁷⁵ as the initiation codon.

The molecular weight of the protein, calculated after removal of the signal peptide, is 57.4 kDa, which is greater than the molecular weight determined for the purified extracellular κ -carrageenase, namely 40.0 kDa.

SECTION 3: Comparison of the protein sequences of the κ -carrageenases of *Alteromonas carrageenovora* and *Cytophaga drobachiensis*

The κ -carrageenase of *C. drobachiensis* has a similarity of 36.1% with the κ -carrageenase of *Alteromonas carrageenovora* over the whole of the linear sequence alignment.

This similarity is particularly high between amino acids 117 and 262 (51.8%) (numbering of the κ -carrageenase of *Alteromonas carrageenovora*) (Fig. 3).

As previously, this similarity is substantiated by HCA analysis, which shows an HCA score between the two proteins of 75.4% over said domain of 145 amino acids (Fig. 4).

5

(1) GENERAL INFORMATION:

(A) NAME: LABORATOIRES GOEMAR S.A.
(B) STREET: La Madeleine B.P. 55
(C) CITY: Saint-Malo
(E) COUNTRY: France
(F) POSTAL CODE (ZIP): 35413 Cedex
(G) TELEPHONE: 99 21 53 70
(H) TELEFAX: 99 82 56 17

(ii) TITLE OF INVENTION: Glycolyse hydrolase genes and their use for producing enzymes for the biodegradation of carrageenans

(iii) NUMBER OF SEQUENCES: 8

(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.30 (EPO)

(2) INFORMATION FOR SEQ ID NO: 1:

(A) LENGTH: 2085 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

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(A) NAME/KEY: CDS
(B) LOCATION:join(211..1683, 1880..2083)
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(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

AAGCTTTCCG ATTCTATCAT CGAAGTCATA GGAGTGGGTA AACAAAAAAG CATGAAACTA 60
GCTTTTTAAA ATACAGACTT TCAATATAGG TCGCACACAA TATTAACGAA TAAATAAGCA 120

AATCATATAC	ATAATCATTG	CTTTAAATAT	GTGTTTAATAC	AGATATAAAC	ATAGTATGTT	180
TGTGTTTTTG	GTATCTATCG	GAGTGAAAAC	ATG CGC TTA	TAT TTT AGA	AAG TTG	234
			Met Arg Leu	Tyr Phe Arg	Lys Leu	
			1		5	
TGG TTA ACA AAT TTA TTT TTA GGC GGA GCA CTG GCC TCT TCA GCT GCG						282
Trp Leu Thr Asn Leu Phe Leu Gly Gly Ala Leu Ala Ser Ser Ala Ala						
10			15		20	
ATA GGG GCT GTC TCC CCC AAG ACT TAT AAG GAC GCA GAT TTT TAT GTT						330
Ile Gly Ala Val Ser Pro Lys Thr Tyr Lys Asp Ala Asp Phe Tyr Val						
25			30		35	40
GCC CCT ACT CAA CAA GAT GTT AAC TAT GAT TTA GTT GAT GAT TTT GGC						378
Ala Pro Thr Gln Gln Asp Val Asn Tyr Asp Leu Val Asp Asp Phe Gly						
			45		50	55
GCT AAT GGA AAC GAC ACT AGT GAT GAC AGT AAT GCT TTA CAA AGA GCA						426
Ala Asn Gly Asn Asp Thr Ser Asp Asp Ser Asn Ala Leu Gln Arg Ala						
			60		65	70
ATT AAT GCT ATT AGT AGA AAA CCG AAT GGG GGC ACT TTA CTA ATA CCG						474
Ile Asn Ala Ile Ser Arg Lys Pro Asn Gly Gly Thr Leu Leu Ile Pro						
			75		80	85
AAT GGA ACT TAC CAT TTC CTC GGC ATA CAG ATG AAG TCG AAC GTA CAC						522
Asn Gly Thr Tyr His Phe Leu Gly Ile Gln Met Lys Ser Asn Val His						
			90		95	100
ATC CGT GTT GAG AGT GAC GTG ATA ATC AAG CCA ACG TGG AAT GGG GAT						570
Ile Arg Val Glu Ser Asp Val Ile Ile Lys Pro Thr Trp Asn Gly Asp						
105			110		115	120
GGC AAA AAC CAC CGA CTA TTT GAA GTT GGC GTA AAC AAT ATT GTA AGA						618
Gly Lys Asn His Arg Leu Phe Glu Val Gly Val Asn Asn Ile Val Arg						
			125		130	135
AAC TTC AGC TTT CAA GGG TTA GGA AAC GGT TTT TTG GTG GAT TTT AAA						666
Asn Phe Ser Phe Gln Gly Leu Gly Asn Gly Phe Leu Val Asp Phe Lys						
			140		145	150
GAT TCT CGC GAC AAA AAC TTA GCT GTT TTT AAG TTA GGC GAT GTT AGA						714
Asp Ser Arg Asp Lys Asn Leu Ala Val Phe Lys Leu Gly Asp Val Arg						
			155		160	165

AAT	TAC	AAA	ATT	TCC	AAT	TTT	ACC	ATT	GAT	GAT	AAT	AAA	ACG	ATA	TTT	762
Asn	Tyr	Lys	Ile	Ser	Asn	Phe	Thr	Ile	Asp	Asp	Asn	Lys	Thr	Ile	Phe	
170						175					180					
GCC	TCA	ATT	TTA	GTG	GAC	GTA	ACA	GAA	CGT	AAT	GGG	CGG	TTA	CAT	TGG	810
Ala	Ser	Ile	Leu	Val	Asp	Val	Thr	Glu	Arg	Asn	Gly	Arg	Leu	His	Trp	
185					190					195					200	
TCG	CGT	AAT	GGA	ATT	ATC	GAA	AGA	ATA	AAA	CAA	AAT	AAC	GCT	TTG	TTC	858
Ser	Arg	Asn	Gly	Ile	Ile	Glu	Arg	Ile	Lys	Gln	Asn	Asn	Ala	Leu	Phe	
				205					210					215		
GGC	TAC	GGC	CTT	ATT	CAA	ACC	TAT	GGC	GCA	GAT	AAT	ATT	TTG	TTT	AGG	906
Gly	Tyr	Gly	Leu	Ile	Gln	Thr	Tyr	Gly	Ala	Asp	Asn	Ile	Leu	Phe	Arg	
			220					225					230			
AAC	CTC	CAT	TCG	GAA	GGC	GGA	ATT	GCG	TTA	CGG	ATG	GAA	ACT	GAC	AAC	954
Asn	Leu	His	Ser	Glu	Gly	Gly	Ile	Ala	Leu	Arg	Met	Glu	Thr	Asp	Asn	
		235					240					245				
TTA	CTT	ATG	AAA	AAT	TAT	AAG	CAA	GGC	GGA	ATA	AGA	AAC	ATC	TTT	GCT	1002
Leu	Leu	Met	Lys	Asn	Tyr	Lys	Gln	Gly	Gly	Ile	Arg	Asn	Ile	Phe	Ala	
		250				255					260					
GAT	AAT	ATC	AGA	TGT	AGC	AAA	GGA	CTT	GCG	GCG	GTC	ATG	TTT	GGC	CCA	1050
Asp	Asn	Ile	Arg	Cys	Ser	Lys	Gly	Leu	Ala	Ala	Val	Met	Phe	Gly	Pro	
265					270					275					280	
CAT	TTT	ATG	AAG	AAT	GGA	GAT	GTG	CAA	GTG	ACC	AAT	GTC	AGC	TCA	GTT	1098
His	Phe	Met	Lys	Asn	Gly	Asp	Val	Gln	Val	Thr	Asn	Val	Ser	Ser	Val	
				285					290					295		
AGT	TGC	GGT	TCG	GCT	GTA	CGA	AGT	GAT	AGT	GGA	TTT	GTC	GAA	CTC	TTT	1146
Ser	Cys	Gly	Ser	Ala	Val	Arg	Ser	Asp	Ser	Gly	Phe	Val	Glu	Leu	Phe	
			300					305					310			
AGC	CCG	ACA	GAC	GAA	GTA	CAT	ACG	CGT	CAA	AGT	TGG	AAA	CAA	GCC	GTT	1194
Ser	Pro	Thr	Asp	Glu	Val	His	Thr	Arg	Gln	Ser	Trp	Lys	Gln	Ala	Val	
		315					320					325				
GAA	AGT	AAA	TTG	GGC	CGA	GGG	TGT	GCG	CAA	ACC	CCT	TAT	GCT	AGA	GGT	1242
Glu	Ser	Lys	Leu	Gly	Arg	Gly	Cys	Ala	Gln	Thr	Pro	Tyr	Ala	Arg	Gly	
		330				335					340					

[illegible]

AAA AAT GCT CTT TTA TTT GCA GGC TTT TCG TTA AGT CTA GTT GCA CAG	1945
Lys Asn Ala Leu Leu Phe Ala Gly Phe Ser Leu Ser Leu Val Ala Gln	
500 505 510	
TCA GTT AGT GCA CAA GAA GCA AAA CAG CCT GAA AAA GAA GAA AAA GAT	1993
Ser Val Ser Ala Gln Glu Ala Lys Gln Pro Glu Lys Glu Glu Lys Asp	
515 520 525	
GTT GAG GTG ATT TTG GTA TCG GCA CAA AAG CGT GAG CAA GCG CTT AAA	2041
Val Glu Val Ile Leu Val Ser Ala Gln Lys Arg Glu Gln Ala Leu Lys	
530 535 540 545	
GAA GTG CCT GTA TCA ATT GAA GTT ATT CAA GGC GAC CTT CTA GA	2085
Glu Val Pro Val Ser Ile Glu Val Ile Gln Gly Asp Leu Leu	
550 555	

(2) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 559 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

Met	Arg	Leu	Tyr	Phe	Arg	Lys	Leu	Trp	Leu	Thr	Asn	Leu	Phe	Leu	Gly
1				5				10						15	
Gly	Ala	Leu	Ala	Ser	Ser	Ala	Ala	Ile	Gly	Ala	Val	Ser	Pro	Lys	Thr
		20						25						30	
Tyr	Lys	Asp	Ala	Asp	Phe	Tyr	Val	Ala	Pro	Thr	Gln	Gln	Asp	Val	Asn
		35					40						45		
Tyr	Asp	Leu	Val	Asp	Asp	Phe	Gly	Ala	Asn	Gly	Asn	Asp	Thr	Ser	Asp
		50					55				60				
Asp	Ser	Asn	Ala	Leu	Gln	Arg	Ala	Ile	Asn	Ala	Ile	Ser	Arg	Lys	Pro
		65				70				75					80
Asn	Gly	Gly	Thr	Leu	Ile	Pro	Asn	Gly	Thr	Tyr	His	Phe	Leu	Gly	
			85					90					95		
Ile	Gln	Met	Lys	Ser	Asn	Val	His	Ile	Arg	Val	Glu	Ser	Asp	Val	Ile
		100						105					110		
Ile	Lys	Pro	Thr	Trp	Asn	Gly	Asp	Gly	Lys	Asn	His	Arg	Leu	Phe	Glu
		115				120						125			
Val	Gly	Val	Asn	Asn	Ile	Val	Arg	Asn	Phe	Ser	Phe	Gln	Gly	Leu	Gly
		130				135						140			

TOP SECRET 00228550

Asn	Gly	Phe	Leu	Val	Asp	Phe	Lys	Asp	Ser	Arg	Asp	Lys	Asn	Leu	Ala
145					150					155					160
Val	Phe	Lys	Leu	Gly	Asp	Val	Arg	Asn	Tyr	Lys	Ile	Ser	Asn	Phe	Thr
				165					170						175
Ile	Asp	Asp	Asn	Lys	Thr	Ile	Phe	Ala	Ser	Ile	Leu	Val	Asp	Val	Thr
				180					185						190
Glu	Arg	Asn	Gly	Arg	Leu	His	Trp	Ser	Arg	Asn	Gly	Ile	Ile	Glu	Arg
				195				200							205
Ile	Lys	Gln	Asn	Asn	Ala	Leu	Phe	Gly	Tyr	Gly	Leu	Ile	Gln	Thr	Tyr
						215									220
Gly	Ala	Asp	Asn	Ile	Leu	Phe	Arg	Asn	Leu	His	Ser	Glu	Gly	Gly	Ile
225					230					235					240
Ala	Leu	Arg	Met	Glu	Thr	Asp	Asn	Leu	Leu	Met	Lys	Asn	Tyr	Lys	Gln
				245					250						255
Gly	Gly	Ile	Arg	Asn	Ile	Phe	Ala	Asp	Asn	Ile	Arg	Cys	Ser	Lys	Gly
				260				265							270
Leu	Ala	Ala	Val	Met	Phe	Gly	Pro	His	Phe	Met	Lys	Asn	Gly	Asp	Val
				275				280							285
Gln	Val	Thr	Asn	Val	Ser	Ser	Val	Ser	Cys	Gly	Ser	Ala	Val	Arg	Ser
						295						300			
Asp	Ser	Gly	Phe	Val	Glu	Leu	Phe	Ser	Pro	Thr	Asp	Glu	Val	His	Thr
305					310						315				320
Arg	Gln	Ser	Trp	Lys	Gln	Ala	Val	Glu	Ser	Lys	Leu	Gly	Arg	Gly	Cys
				325					330						335
Ala	Gln	Thr	Pro	Tyr	Ala	Arg	Gly	Asn	Gly	Gly	Thr	Arg	Trp	Ala	Ala
				340				345							350
Arg	Val	Thr	Gln	Lys	Asp	Ala	Cys	Leu	Asp	Lys	Ala	Lys	Leu	Glu	Tyr
				355				360					365		
Gly	Ile	Glu	Pro	Gly	Ser	Phe	Gly	Thr	Val	Lys	Val	Phe	Asp	Val	Thr
						375						380			
Ala	Arg	Phe	Gly	Tyr	Asn	Ala	Asp	Leu	Lys	Gln	Asp	Gln	Leu	Asp	Tyr
385					390					395					400
Phe	Ser	Thr	Ser	Asn	Pro	Met	Cys	Lys	Arg	Val	Cys	Leu	Pro	Thr	Lys
				405					410						415
Glu	Gln	Trp	Ser	Lys	Gln	Gly	Gln	Ile	Tyr	Ile	Gly	Pro	Ser	Leu	Ala
				420				425							430
Ala	Val	Ile	Asp	Thr	Thr	Pro	Glu	Thr	Ser	Lys	Tyr	Asp	Tyr	Asp	Val
				435				440					445		
Lys	Thr	Phe	Asn	Val	Lys	Arg	Ile	Asn	Phe	Pro	Val	Asn	Ser	His	Lys
						455					460				
Thr	Ile	Asp	Thr	Asn	Thr	Glu	Ser	Ser	Arg	Val	Cys	Asn	Tyr	Tyr	Gly
465					470					475					480
Met	Ser	Glu	Cys	Ser	Ser	Ser	Arg	Trp	Glu	Arg	Met	Lys	Gly	Val	Ser
				485					490						495
Thr	Lys	Asn	Ala	Leu											

(2) INFORMATION FOR SEQ ID NO: 3:

(A) LENGTH: 1997 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(iii) HYPOTHETICAL: NO

```
(A) NAME/KEY: CDS
(B) LOCATION:join(333..1805, 1866..1997)
```

CCCTAAAAAC	TATTCTTCAT	ACCCTTTGAT	GTATACGTTT	AAACTATAGG	GAGTTAATCT	60
GGTTTTGGTG	CAATTCTAGT	TTAATAAATG	AAGCCTTCTT	TTTGGACTTA	CATTTTATTA	120
ACCTCTTGAA	TTCTTGGGGC	TTGCTAATTA	TAAAATACTT	AATATCAGGT	GGTTGTGTAA	180
AAGAGGTGGA	AGGGTATAGG	ACCGTTACTT	ATAATTGGCC	CCTGTCGGAA	GGGGGGTTAA	240
AGGTAAATA	GTGTTTAAGT	GTATTAATTA	ACTTCTATAT	AAGTAGGAAA	ATACACTATA	300
TATTGCGACA	TTATTAACCT	TAAATTCTTA	CA ATG AAA	TTA CAA TTT	AAA CCT	353
			Met Lys	Leu Gln Phe	Lys Pro	
			1		5	

GTG ACG GAA AAC GAT ACC TCC GAA ATT TCG GAA GTT CCA ACT GAA TTG 449
Val Thr Glu Asn Asp Thr Ser Glu Ile Ser Glu Val Pro Thr Glu Leu
25 30 35

AGG GCC GCG GCT TCT TCA TTT TAT ACC CCA CCG GGT CAG AAT GTA CGG 497
Arg Ala Ala Ala Ser Ser Phe Tyr Thr Pro Pro Gly Gln Asn Val Arg
40 45 50 55

GCC	AAT	AAA	AAA	AAC	CTG	GTC	ACG	GAT	TAC	GGT	GTT	AAC	CAC	AAT	GAT	545
Ala	Asn	Lys	Lys	Asn	Leu	Val	Thr	Asp	Tyr	Gly	Val	Asn	His	Asn	Asp	
				60					65					70		
CAG	AAC	GAT	GAT	AGT	AGC	AAA	TTA	AAC	CTG	GCT	ATC	AAA	GAT	TTA	TCG	593
Gln	Asn	Asp	Asp	Ser	Ser	Lys	Leu	Asn	Leu	Ala	Ile	Lys	Asp	Leu	Ser	
				75				80					85			
GAT	ACC	GGT	GGT	ATA	CTG	ACC	CTT	CCT	AAG	GGA	AAG	TAC	TAT	TTG	ACC	641
Asp	Thr	Gly	Gly	Ile	Leu	Thr	Leu	Pro	Lys	Gly	Lys	Tyr	Tyr	Leu	Thr	
		90					95					100				
AAA	ATT	AGA	ATG	CGC	TCT	AAT	GTA	CAT	CTT	GAA	ATA	GAA	AAG	GGA	ACG	689
Lys	Ile	Arg	Met	Arg	Ser	Asn	Val	His	Leu	Glu	Ile	Glu	Lys	Gly	Thr	
	105					110				115						
GTA	ATC	TAT	CCG	ACC	AAG	GGG	TTG	ACT	CCT	GCG	AAG	AAT	CAC	AGA	ATT	737
Val	Ile	Tyr	Pro	Thr	Lys	Gly	Leu	Thr	Pro	Ala	Lys	Asn	His	Arg	Ile	
120					125					130					135	
TTT	GAT	TTT	GCC	AGT	AAA	ACA	GAG	GAA	AAA	ATA	GAA	AAC	GCC	AGT	ATA	785
Phe	Asp	Phe	Ala	Ser	Lys	Thr	Glu	Glu	Lys	Ile	Glu	Asn	Ala	Ser	Ile	
				140					145					150		
GTG	GGT	AAA	GGA	GGT	AAG	TTT	ATA	GTA	GAC	CTA	AGA	GGC	AAC	AGT	TCT	833
Val	Gly	Lys	Gly	Gly	Lys	Phe	Ile	Val	Asp	Leu	Arg	Gly	Asn	Ser	Ser	
			155					160					165			
AAA	AAC	CAA	ATT	GTA	GCC	GAT	GTT	GGT	AAC	GTA	ACC	AAC	TTT	AAA	ATA	881
Lys	Asn	Gln	Ile	Val	Ala	Asp	Val	Gly	Asn	Val	Thr	Asn	Phe	Lys	Ile	
		170					175					180				
TCG	AAT	TTT	ACG	ATC	AAG	GAT	GAA	AAA	ACC	ATC	TTT	GCT	TCG	ATA	TTG	929
Ser	Asn	Phe	Thr	Ile	Lys	Asp	Glu	Lys	Thr	Ile	Phe	Ala	Ser	Ile	Leu	
	185					190					195					
GTA	AGC	TTT	ACG	GAT	AAG	GCA	GGC	AAT	GCT	TGG	CCA	CAT	AAA	GGT	ATT	977
Val	Ser	Phe	Thr	Asp	Lys	Ala	Gly	Asn	Ala	Trp	Pro	His	Lys	Gly	Ile	
200					205					210					215	
ATT	GAG	AAT	ATA	GAC	CAG	GCG	AAT	GCC	CAT	ACG	GGA	TAT	GGC	CTC	ATA	1025
Ile	Glu	Asn	Ile	Asp	Gln	Ala	Asn	Ala	His	Thr	Gly	Tyr	Gly	Leu	Ile	
				220					225					230		

CAG	GCG	TAC	GCG	GCA	GAT	AAC	ATT	CTG	TTC	AAC	AAT	CTA	AGT	TGT	ACG
Gln	Ala	Tyr	Ala	Ala	Asp	Asn	Ile	Leu	Phe	Asn	Asn	Leu	Ser	Cys	Thr
			235					240					245		
GGC	GGG	GTA	ACC	TTG	CGT	TTA	GAA	ACC	GAC	AAC	CTC	GCT	ATG	AAA	ACC
Gly	Gly	Val	Thr	Leu	Arg	Leu	Glu	Thr	Asp	Asn	Leu	Ala	Met	Lys	Thr
			250					255					260		
GCT	AAA	AAA	GGG	GGG	GTA	AGG	GAT	ATT	TTT	GCC	ACA	AAG	ATC	AAG	AAT
Ala	Lys	Lys	Gly	Gly	Val	Arg	Asp	Ile	Phe	Ala	Thr	Lys	Ile	Lys	Asn
			265					270					275		
ACC	AAT	GGC	TTG	ACC	CCG	GTA	ATG	TTC	TCT	CCC	CAT	TTT	ATG	GAA	AAC
Thr	Asn	Gly	Leu	Thr	Pro	Val	Met	Phe	Ser	Pro	His	Phe	Met	Glu	Asn
						285					290				295
GGT	AAA	GTG	ACC	ATA	GAT	GAT	GTA	ACC	GCC	ATC	GGT	TGT	GCA	TAT	GCC
Gly	Lys	Val	Thr	Ile	Asp	Asp	Val	Thr	Ala	Ile	Gly	Cys	Ala	Tyr	Ala
				300					305					310	
GTA	CGT	GTA	GAG	CAC	GGT	TTT	ATA	GAG	ATT	TTC	GAT	AAG	GGG	AAT	AGG
Val	Arg	Val	Glu	His	Gly	Phe	Ile	Glu	Ile	Phe	Asp	Lys	Gly	Asn	Arg
			315					320					325		
GCA	AGT	GCC	GAC	GCT	TTC	AAG	AAC	TAT	ATT	GAA	GGT	ATT	CTA	GGA	GCT
Ala	Ser	Ala	Asp	Ala	Phe	Lys	Asn	Tyr	Ile	Glu	Gly	Ile	Leu	Gly	Ala
			330					335					340		
GGC	TCG	GTA	GAA	GTC	GTG	TAC	AAA	CGT	AAT	AAC	GGA	AGA	ACA	TGG	GCG
Gly	Ser	Val	Glu	Val	Val	Tyr	Lys	Arg	Asn	Asn	Gly	Arg	Thr	Trp	Ala
			345					350				355			
GCA	CGT	ATC	GCA	AAC	GAC	TTT	AAC	GAA	GCG	GCG	TAT	AAC	CAC	TCC	AAT
Ala	Arg	Ile	Ala	Asn	Asp	Phe	Asn	Glu	Ala	Ala	Tyr	Asn	His	Ser	Asn
			360				365			370					375
CCT	GCC	GTT	AGC	GGA	ATC	AAA	CCA	GGG	AAA	TTC	GCC	ACA	TCT	AAG	GTA
Pro	Ala	Val	Ser	Gly	Ile	Lys	Pro	Gly	Lys	Phe	Ala	Thr	Ser	Lys	Val
				380					385					390	
ACC	AAT	GTT	AAG	GCA	ACC	TAT	AAG	GGT	ACT	GGC	GCC	AAA	CTC	AAG	CAG
Thr	Asn	Val	Lys	Ala	Thr	Tyr	Lys	Gly	Thr	Gly	Ala	Lys	Leu	Lys	Gln
			395					400					405		

GCA TTC TTA TCC TAT TTA CCC TGT TCG GAA CGT TCT AAG GTT TGT CGG 1601
 Ala Phe Leu Ser Tyr Leu Pro Cys Ser Glu Arg Ser Lys Val Cys Arg
 410 415 420

CCA GGT CCA GAT GGG TTC GAG TAT AAC GGA CCC TCC TTG GGA GTT ACC 1649
 Pro Gly Pro Asp Gly Phe Glu Tyr Asn Gly Pro Ser Leu Gly Val Thr
 425 430 435

ATC GAT AAC ACG AAA AGG GAC AAC AGC CTT GGC AAT TAT AAC GTC AAT 1697
 Ile Asp Asn Thr Lys Arg Asp Asn Ser Leu Gly Asn Tyr Asn Val Asn
 440 445 450 455

GTA AGC ACC TCC AGT GTT CAG GGC TTT CCC AAT AAT TAC GTT TTA AAC 1745
 Val Ser Thr Ser Ser Val Gln Gly Phe Pro Asn Asn Tyr Val Leu Asn
 460 465 470

GTA AAG TAT AAT ACC CCT AAA GTA TGT AAC CAA AAT CTA GGT AGT ATT 1793
 Val Lys Tyr Asn Thr Pro Lys Val Cys Asn Gln Asn Leu Gly Ser Ile
 475 480 485

ACT TCG TGT AAC TGATCACGAA ACAATTTGTA AATAAAAAGC AGCTGTCCCT 1845
 Thr Ser Cys Asn
 490

TATTACGGGC GGCTGCTTTT ATG TCT TTA AGC CAT GTC GTG ATT TAT TGG 1895
 Met Ser Leu Ser His Val Val Ile Tyr Trp
 495 500

CGA CTT TTG ATA AAG GCT TGG ATT TCT TCC GGG GTA AAT ATC GGA TTG 1943
 Arg Leu Leu Ile Lys Ala Trp Ile Ser Ser Gly Val Asn Ile Gly Leu
 505 510 515

GCC CCT TCC CTA CCG GCT ACC ATA GCT CTA TGC TCC TAT GCA CAG GCG 1991
 Ala Pro Ser Leu Pro Ala Thr Ile Ala Leu Cys Ser Tyr Ala Gln Ala
 520 525 530

AAA TCT 1997
 Lys Ser
 535

(2) INFORMATION FOR SEQ ID NO: 4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 535 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

```

Met Lys Leu Gln Phe Lys Pro Val Tyr Leu Ala Ser Ile Ala Ile Met
 1           5           10           15
Ala Ile Gly Cys Thr Lys Glu Val Thr Glu Asn Asp Thr Ser Glu Ile
          20           25           30
Ser Glu Val Pro Thr Glu Leu Arg Ala Ala Ala Ser Ser Phe Tyr Thr
          35           40           45
Pro Pro Gly Gln Asn Val Arg Ala Asn Lys Lys Asn Leu Val Thr Asp
          50           55           60
Tyr Gly Val Asn His Asn Asp Gln Asn Asp Asp Ser Ser Lys Leu Asn
          65           70           75           80
Leu Ala Ile Lys Asp Leu Ser Asp Thr Gly Gly Ile Leu Thr Leu Pro
          85           90           95
Lys Gly Lys Tyr Tyr Leu Thr Lys Ile Arg Met Arg Ser Asn Val His
          100          105          110
Leu Glu Ile Glu Lys Gly Thr Val Ile Tyr Pro Thr Lys Gly Leu Thr
          115          120          125
Pro Ala Lys Asn His Arg Ile Phe Asp Phe Ala Ser Lys Thr Glu Glu
          130          135          140
Lys Ile Glu Asn Ala Ser Ile Val Gly Lys Gly Gly Lys Phe Ile Val
          145          150          155          160
Asp Leu Arg Gly Asn Ser Ser Lys Asn Gln Ile Val Ala Asp Val Gly
          165          170          175
Asn Val Thr Asn Phe Lys Ile Ser Asn Phe Thr Ile Lys Asp Glu Lys
          180          185          190
Thr Ile Phe Ala Ser Ile Leu Val Ser Phe Thr Asp Lys Ala Gly Asn
          195          200          205
Ala Trp Pro His Lys Gly Ile Ile Glu Asn Ile Asp Gln Ala Asn Ala
          210          215          220
His Thr Gly Tyr Gly Leu Ile Gln Ala Tyr Ala Ala Asp Asn Ile Leu
          225          230          235          240
Phe Asn Asn Leu Ser Cys Thr Gly Gly Val Thr Leu Arg Leu Glu Thr
          245          250          255
Asp Asn Leu Ala Met Lys Thr Ala Lys Lys Gly Gly Val Arg Asp Ile
          260          265          270
Phe Ala Thr Lys Ile Lys Asn Thr Asn Gly Leu Thr Pro Val Met Phe
          275          280          285
Ser Pro His Phe Met Glu Asn Gly Lys Val Thr Ile Asp Asp Val Thr
          290          295          300
Ala Ile Gly Cys Ala Tyr Ala Val Arg Val Glu His Gly Phe Ile Glu
          305          310          315          320

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00988200 44504

(2) INFORMATION FOR SEQ ID NO: 5:

(A) LENGTH: 2180 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(iii) HYPOTHETICAL: NO

```
(A) NAME/KEY: CDS
(B) LOCATION:join(1..498, 741..1931, 2009..2179)
```

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

[illegible]

648
708
761

809

857

905

953

1001

1049

1097

1145

1193

1241

GAA	CTT	ACT	CAA	AAA	AGT	GCA	GTG	AGA	GAG	TCT	GAT	CAT	GAC	TTA	CAC	1289
Glu	Leu	Thr	Gln	Lys	Ser	Ala	Val	Arg	Glu	Ser	Asp	His	Asp	Leu	His	
335						340					345					
AAT	ATT	GTA	GTA	AAA	AAT	GGA	AAA	CCA	ACA	TGG	ATG	CGT	CCA	GGG	TCT	1337
Asn	Ile	Val	Val	Lys	Asn	Gly	Lys	Pro	Thr	Trp	Met	Arg	Pro	Gly	Ser	
350					355					360					365	
TTT	CCG	CAG	ACA	AAT	CAT	AAC	GGA	TAC	CAT	CTA	CCT	TTC	GAT	CCT	CGA	1385
Phe	Pro	Gln	Thr	Asn	His	Asn	Gly	Tyr	His	Leu	Pro	Phe	Asp	Pro	Arg	
				370					375					380		
AAT	GAC	TTT	CAC	ACC	TAT	GGT	GTC	AAT	GTA	ACT	AAA	GAC	AAG	ATC	ACT	1433
Asn	Asp	Phe	His	Thr	Tyr	Gly	Val	Asn	Val	Thr	Lys	Asp	Lys	Ile	Thr	
			385					390					395			
TGG	TAC	GTA	GAT	GGT	GAA	ATT	GTG	GGC	GAA	AAG	GAT	AAC	TTA	TAC	TGG	1481
Trp	Tyr	Val	Asp	Gly	Glu	Ile	Val	Gly	Glu	Lys	Asp	Asn	Leu	Tyr	Trp	
		400					405					410				
CAT	CGT	CAA	ATG	AAT	CTC	ACA	TTA	TCA	CAA	GGC	TTA	CGC	GCG	CCG	CAT	1529
His	Arg	Gln	Met	Asn	Leu	Thr	Leu	Ser	Gln	Gly	Leu	Arg	Ala	Pro	His	
	415					420					425					
ACA	CAA	TGG	AAA	TGT	AAT	CAA	TTT	TAC	CCA	TCA	GCG	AAT	AAA	TCA	GCA	1577
Thr	Gln	Trp	Lys	Cys	Asn	Gln	Phe	Tyr	Pro	Ser	Ala	Asn	Lys	Ser	Ala	
430					435					440					445	
GAA	GGC	TTC	CCA	ACA	TCA	ATG	GAA	GTT	GAT	TAT	GTA	AGA	ACG	TGG	GTA	1625
Glu	Gly	Phe	Pro	Thr	Ser	Met	Glu	Val	Asp	Tyr	Val	Arg	Thr	Trp	Val	
				450					455					460		
AAG	GTG	GGC	AAT	AAC	AAC	TCT	GCT	CCA	GGC	GAG	GGG	CAG	TCA	TGT	CCT	1673
Lys	Val	Gly	Asn	Asn	Asn	Ser	Ala	Pro	Gly	Glu	Gly	Gln	Ser	Cys	Pro	
			465				470						475			
AAC	ACG	TTT	GTA	GCT	GTC	AAT	AGT	GTT	CAA	CTA	AGC	GCA	GCA	AAA	CAA	1721
Asn	Thr	Phe	Val	Ala	Val	Asn	Ser	Val	Gln	Leu	Ser	Ala	Ala	Lys	Gln	
		480					485					490				
ACA	CTT	CGA	AAG	GGC	CAA	TCT	ACA	ACG	CTA	GAA	AGC	ACA	GTT	CTT	CCA	1769
Thr	Leu	Arg	Lys	Gly	Gln	Ser	Thr	Thr	Leu	Glu	Ser	Thr	Val	Leu	Pro	
	495					500					505					

TCA AAA GGA AAG CTT GAT C 2180
Ser Lys Gly Lys Leu Asp
615 620

(ii) MOLECULE TYPE: protein

Asp	His	Ile	Ile	Pro	Leu	Gln	Ile	Lys	Asn	Ser	Gln	Asp	Ser	Gln	Ile
1				5					10						15
Ile	Ser	Phe	Phe	Lys	Ala	Asp	Lys	Gly	Ser	Val	Ser	Arg	Gln	Val	His
			20					25					30		
Pro	Pro	Trp	Pro	Val	Pro	Cys	Lys	Ser	Lys	Leu	Gln	Glu	Gln	Asp	Ser
		35					40					45			
Ser	Glu	Ser	Lys	Glu	Ser	Lys	Ala	Glu	Gln	Val	Lys	Ile	Asn	Asn	Cys
	50					55					60				
Val	Val	Gln	Asn	Ala	Met	Leu	Tyr	Ile	Glu	Asn	Asn	Tyr	Phe	Asn	Asp
65					70					75					80
Ile	Asn	Ile	Asp	Thr	Val	Ala	Phe	Ser	Val	Gly	Val	Ser	Arg	Ser	Tyr
				85					90					95	
Leu	Val	Lys	Gln	Phe	Lys	Leu	Ala	Thr	Asn	Lys	Thr	Ile	Asn	Asn	Arg
			100					105					110		
Ile	Ile	Glu	Val	Arg	Ile	Glu	Gln	Ala	Lys	Lys	Val	Leu	Leu	Lys	Lys
		115					120					125			
Ser	Val	Thr	Glu	Thr	Ala	Tyr	Glu	Val	Gly	Phe	Asn	Asn	Ser	Asn	Tyr
	130					135					140				
Phe	Ala	Thr	Val	Phe	Lys	Lys	Arg	Thr	Asn	Tyr	Thr	Pro	Lys	Gln	Phe
145					150					155					160
Lys	Arg	Thr	Phe	Ser	Ser	Met	Lys	Pro	Ile	Ser	Ile	Val	Ala	Phe	Pro
				165					170					175	
Ile	Pro	Ala	Ile	Ser	Met	Leu	Leu	Leu	Ser	Ala	Val	Ser	Gln	Ala	Ala
			180					185					190		
Ser	Met	Gln	Pro	Pro	Ile	Ala	Lys	Pro	Gly	Glu	Thr	Trp	Ile	Leu	Gln
		195					200					205			
Ala	Lys	Arg	Ser	Asp	Glu	Phe	Asn	Val	Lys	Asp	Ala	Thr	Lys	Trp	Asn
	210					215					220				
Phe	Gln	Thr	Glu	Asn	Tyr	Gly	Val	Trp	Ser	Trp	Lys	Asn	Glu	Asn	Ala
225					230					235					240
Thr	Val	Ser	Asn	Gly	Lys	Leu	Lys	Leu	Thr	Thr	Lys	Arg	Glu	Ser	His
				245					250					255	
Gln	Arg	Thr	Phe	Trp	Asp	Gly	Cys	Asn	Gln	Gln	Gln	Val	Ala	Asn	Tyr
			260					265					270		
Pro	Leu	Tyr	Tyr	Thr	Ser	Gly	Val	Ala	Lys	Ser	Arg	Ala	Thr	Gly	Asn
	275					280						285			
Tyr	Gly	Tyr	Tyr	Glu	Ala	Arg	Ile	Lys	Gly	Ala	Ser	Thr	Phe	Pro	Gly
	290					295					300				
Val	Ser	Pro	Ala	Phe	Trp	Met	Tyr	Ser	Thr	Ile	Asp	Arg	Ser	Leu	Thr
305					310					315					320
Lys	Glu	Gly	Asp	Val	Gln	Tyr	Ser	Glu	Ile	Asp	Val	Val	Glu	Leu	Thr
				325					330					335	
Gln	Lys	Ser	Ala	Val	Arg	Glu	Ser	Asp	His	Asp	Leu	His	Asn	Ile	Val
			340					345					350		

Val Lys Asn Gly Lys Pro Thr Trp Met Arg Pro Gly Ser Phe Pro Gln
 355 360 365
 Thr Asn His Asn Gly Tyr His Leu Pro Phe Asp Pro Arg Asn Asp Phe
 370 375 380
 His Thr Tyr Gly Val Asn Val Thr Lys Asp Lys Ile Thr Trp Tyr Val
 385 390 395 400
 Asp Gly Glu Ile Val Gly Glu Lys Asp Asn Leu Tyr Trp His Arg Gln
 405 410 415
 Met Asn Leu Thr Leu Ser Gln Gly Leu Arg Ala Pro His Thr Gln Trp
 420 425 430
 Lys Cys Asn Gln Phe Tyr Pro Ser Ala Asn Lys Ser Ala Glu Gly Phe
 435 440 445
 Pro Thr Ser Met Glu Val Asp Tyr Val Arg Thr Trp Val Lys Val Gly
 450 455 460
 Asn Asn Asn Ser Ala Pro Gly Glu Gly Gln Ser Cys Pro Asn Thr Phe
 465 470 475 480
 Val Ala Val Asn Ser Val Gln Leu Ser Ala Ala Lys Gln Thr Leu Arg
 485 490 495
 Lys Gly Gln Ser Thr Thr Leu Glu Ser Thr Val Leu Pro Asn Cys Ala
 500 505 510
 Thr Asn Lys Lys Val Ile Tyr Ser Ser Ser Asn Lys Asn Val Ala Thr
 515 520 525
 Val Asn Ser Ala Gly Val Val Lys Ala Lys Asn Lys Gly Thr Ala Thr
 530 535 540
 Ile Thr Val Lys Thr Lys Asn Lys Gly Lys Ile Asp Lys Leu Thr Ile
 545 550 555 560
 Ala Val Asn Met Lys Lys Val Asn Leu Ser Ser Lys Trp Ile Ile Ser
 565 570 575
 Ile Ser Leu Leu Ile Ile Cys Asp Tyr Val Tyr Leu Ile Arg Thr Asn
 580 585 590
 Val Asn Glu Gln Ala Asn Ala Glu Ala Thr Ala His Met His Tyr Lys
 595 600 605
 Ile Asn Asn Thr Lys His Ser Lys Gly Lys Leu Asp
 610 615 620

(2) INFORMATION FOR SEQ ID NO: 7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2600 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION:875..2509

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

GCCTCCGTAT TCGACAATGT TGTACGATGC TTGGCGATTC GGACTCTGTT TAAGCACTCG 60
 ATTTTCGTAAA GGCACATATCC ACTCATTTCAT TCCGACTCAA TATTCTTTTC GACAAATGCA 120
 ACCGGTTCCA TTGAAAAGGC CCTAAAAATA CAGCTTTCCC GCCCCCATC GTAGAAGGTT 180
 CCAATATGCT TCAACCCCTT TTTCAGCCTT ACTTCAGGGG TATTACTTTC ATGCCTAGGG 240
 CCGCAAATAC ATTCGCTTGG ACCCAGTCAC CTATATAATT GAATACGGAA CTACCCATGG 300
 CTTCTTCCC TTTGGGAACC TATGGTACAG ACTTGCTTTT TTAAACCGG TTAATTCAGC 360
 TAATTCGCCA AGCTGGTTCC TTCATAACCT TTGGCCCGAA ACACCTTGCA AGCACATAAA 420
 TCTTATCCAA TATTTTGCGG TCTCATGGGA CAAATCTATA ACAAACATTC AATTTTACCA 480
 AACGTTCCGT AATAAATCTA GTCAAAAACG GGGTCCGATT CATTTTAGAA GAAAGGTAAA 540
 GCCCCAAAA GAGCGGTTTA CTTGAAGATA TGATTTATAA AACACAATAA GTGACAAAGG 600
 AAGATCATGG CTATAATTAG TTGAAAAAAC AGGGCTTACC ATGACATGGA GCTTTATTGA 660
 AAACAGATGT CCAACAAGAA TAAAGGAGGG CCGTTCGACC GCGACGTTTA AATAAAAACA 720
 TATTCCATAT CAAAATTTAA TTAAGGTTCT TTCCTACAGT ATTTATAAGA AATTACTAAA 780
 ATTAGTTAGG ATAATACTAC AAAATGGTAA AATTGGATTA CTCAGATTGA ACCATAGCCT 840
 CTACTTTAGT CGGCTAACAA AAACAATTAT AGTA ATG AAA AAA CCA AAT TTT 892
 Met Lys Lys Pro Asn Phe
 1 5
 TAT GGC AAG ATG GGT AGA ACT GCA CTT TCA AGT CTT TTC TAC CTC TTT 940
 Tyr Gly Lys Met Gly Arg Thr Ala Leu Ser Ser Leu Phe Tyr Leu Phe
 10 15 20
 TTC CTA GGC CTT GTG TAT GGG CAA CAA CCT ACG AAG ACT TCA AAT CCG 988
 Phe Leu Gly Leu Val Tyr Gly Gln Gln Pro Thr Lys Thr Ser Asn Pro
 25 30 35
 AAC GAT CAG TGG ACC ATC AAA TGG AGT GCT TCG GAC GAA TTC AAC AAA 1036
 Asn Asp Gln Trp Thr Ile Lys Trp Ser Ala Ser Asp Glu Phe Asn Lys
 40 45 50
 AAT GAC CCC GAC TGG GCA AAA TGG ATC AAG ACA GGA AAC CTT CCG AAT 1084
 Asn Asp Pro Asp Trp Ala Lys Trp Ile Lys Thr Gly Asn Leu Pro Asn
 55 60 65 70
 ACA TCG GCA TGG AAA TGG AAC AAT CAA AAA AAC GTA AAG ATT TCC AAC 1132
 Thr Ser Ala Trp Lys Trp Asn Asn Gln Lys Asn Val Lys Ile Ser Asn
 75 80 85

09988200-11001
 T0011 00288660

GGA	ATT	GCG	GAA	CTA	ACG	ATG	AGG	CAT	AAC	GCC	AAT	AAT	ACC	CCA	CCT	1180
Gly	Ile	Ala	Glu	Leu	Thr	Met	Arg	His	Asn	Ala	Asn	Asn	Thr	Pro	Pro	
			90					95					100			
GAC	GGA	GGA	ACC	TAT	TTC	ACC	TCT	GGG	ATA	TTT	AAG	TCG	TAC	CAA	AAA	1228
Asp	Gly	Gly	Thr	Tyr	Phe	Thr	Ser	Gly	Ile	Phe	Lys	Ser	Tyr	Gln	Lys	
			105				110					115				
TTT	ACG	TAT	GGA	TAC	TTT	GAG	GCC	AAA	ATC	CAA	GGA	GCG	GAT	ATA	GGT	1276
Phe	Thr	Tyr	Gly	Tyr	Phe	Glu	Ala	Lys	Ile	Gln	Gly	Ala	Asp	Ile	Gly	
			120			125					130					
GAA	GGC	GTA	TGC	CCA	TCG	TTT	TGG	CTT	TAT	AGT	GAT	TTC	GAC	TAT	TCC	1324
Glu	Gly	Val	Cys	Pro	Ser	Phe	Trp	Leu	Tyr	Ser	Asp	Phe	Asp	Tyr	Ser	
135					140					145					150	
GTA	GCC	AAT	GGG	GAA	ACG	GTA	TAC	AGT	GAA	ATA	GAT	GTA	GTT	GAA	CTA	1372
Val	Ala	Asn	Gly	Glu	Thr	Val	Tyr	Ser	Glu	Ile	Asp	Val	Val	Glu	Leu	
			155					160					165			
CAA	CAA	TTC	GAT	TGG	TAT	GAA	GGC	CAT	CAG	GAC	GAC	ATT	TAC	GAC	ATG	1420
Gln	Gln	Phe	Asp	Trp	Tyr	Glu	Gly	His	Gln	Asp	Asp	Ile	Tyr	Asp	Met	
			170				175						180			
GAC	TTA	AAT	CTA	CAC	GCC	GTT	GTC	AAA	GAA	AAC	GGA	CAG	GGG	GTT	TGG	1468
Asp	Leu	Asn	Leu	His	Ala	Val	Val	Lys	Glu	Asn	Gly	Gln	Gly	Val	Trp	
			185				190					195				
AAA	AGG	CCA	AAA	ATG	TAC	CCT	CAA	GAA	CAG	TTG	AAC	AAA	TGG	AGA	GCC	1516
Lys	Arg	Pro	Lys	Met	Tyr	Pro	Gln	Glu	Gln	Leu	Asn	Lys	Trp	Arg	Ala	
		200				205					210					
ATG	GAC	CCG	AGT	AAA	GAC	TTT	CAT	ATC	TAT	GGT	TGT	GAA	GTG	AAC	CAG	1564
Met	Asp	Pro	Ser	Lys	Asp	Phe	His	Ile	Tyr	Gly	Cys	Glu	Val	Asn	Gln	
215				220						225					230	
AAC	GAA	ATC	ATA	TGG	TAT	GTT	GAC	GGT	GTC	GAG	GTT	GCC	CGA	AAA	CCA	1612
Asn	Glu	Ile	Ile	Trp	Tyr	Val	Asp	Gly	Val	Glu	Val	Ala	Arg	Lys	Pro	
			235					240					245			
AAT	AAA	TAT	TGG	CAT	CGC	CCC	ATG	AAC	GTT	ACC	CTT	TCA	TTG	GGA	CTC	1660
Asn	Lys	Tyr	Trp	His	Arg	Pro	Met	Asn	Val	Thr	Leu	Ser	Leu	Gly	Leu	
			250				255					260				

2188

(2) INFORMATION FOR SEQ ID NO: 8:

(A) LENGTH: 545 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

Met	Lys	Lys	Pro	Asn	Phe	Tyr	Gly	Lys	Met	Gly	Arg	Thr	Ala	Leu	Ser
1				5					10					15	
Ser	Leu	Phe	Tyr	Leu	Phe	Phe	Leu	Gly	Leu	Val	Tyr	Gly	Gln	Gln	Pro
			20					25					30		

Thr Lys Thr Ser Asn Pro Asn Asp Gln Trp Thr Ile Lys Trp Ser Ala
 35 40 45
 Ser Asp Glu Phe Asn Lys Asn Asp Pro Asp Trp Ala Lys Trp Ile Lys
 50 55 60
 Thr Gly Asn Leu Pro Asn Thr Ser Ala Trp Lys Trp Asn Asn Gln Lys
 65 70 75 80
 Asn Val Lys Ile Ser Asn Gly Ile Ala Glu Leu Thr Met Arg His Asn
 85 90 95
 Ala Asn Asn Thr Pro Pro Asp Gly Gly Thr Tyr Phe Thr Ser Gly Ile
 100 105 110
 Phe Lys Ser Tyr Gln Lys Phe Thr Tyr Gly Tyr Phe Glu Ala Lys Ile
 115 120 125
 Gln Gly Ala Asp Ile Gly Glu Gly Val Cys Pro Ser Phe Trp Leu Tyr
 130 135 140
 Ser Asp Phe Asp Tyr Ser Val Ala Asn Gly Glu Thr Val Tyr Ser Glu
 145 150 155 160
 Ile Asp Val Val Glu Leu Gln Gln Phe Asp Trp Tyr Glu Gly His Gln
 165 170 175
 Asp Asp Ile Tyr Asp Met Asp Leu Asn Leu His Ala Val Val Lys Glu
 180 185 190
 Asn Gly Gln Gly Val Trp Lys Arg Pro Lys Met Tyr Pro Gln Glu Gln
 195 200 205
 Leu Asn Lys Trp Arg Ala Met Asp Pro Ser Lys Asp Phe His Ile Tyr
 210 215 220
 Gly Cys Glu Val Asn Gln Asn Glu Ile Ile Trp Tyr Val Asp Gly Val
 225 230 235 240
 Glu Val Ala Arg Lys Pro Asn Lys Tyr Trp His Arg Pro Met Asn Val
 245 250 255
 Thr Leu Ser Leu Gly Leu Arg Lys Pro Phe Val Lys Phe Phe Asp Asn
 260 265 270
 Lys Asn Asn Ala Ile Asn Pro Glu Thr Asp Ala Lys Ala Arg Glu Lys
 275 280 285
 Leu Ser Asp Ile Pro Thr Ser Met Tyr Val Asp Tyr Val Arg Val Trp
 290 295 300
 Glu Lys Ser Ala Gly Asn Thr Thr Asn Pro Pro Thr Ser Glu Val Gly
 305 310 315 320
 Thr Leu Lys Thr Lys Gly Ser Lys Leu Val Ile Asp His Trp Asp Ala
 325 330 335
 Ser Thr Gly Thr Ile Ser Ala Val Ser Asn Asn Thr Lys Thr Gly Gln
 340 345 350
 Tyr Ala Gly Ser Val Asn Asn Ala Ser Ile Ala Gln Ile Val Thr Leu
 355 360 365
 Lys Ala Asn Thr Ser Tyr Lys Val Ser Ala Phe Gly Lys Ala Ser Ser
 370 375 380
 Pro Gly Thr Ser Ala Tyr Leu Gly Ile Ser Lys Ala Ser Asn Asn Glu
 385 390 395 400

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